Occurrence of *Anisakis* spp. in horse mackerel (*Trachurus trachurus* L.) from the North Atlantic Moroccan coasts

Lamyae Azbaid^{1*}, Aziz Lamtai¹, El Mostafa Talbaoui¹, Fatima Chidi²

¹Institut National de Recherche Halieutique (INRH). Centre Régional de Tanger, BP 5268, Tanger 90 040 Morocco ²Faculté des Sciences et Techniques de Tanger. Université Abdelmalek Essaâdi. BP 416 90 000. Tanger -Morocco ^{*}Corresponding autor E-mail: lazbaid@gmail.com

Abstract

The Anisakidae (Nematoda) have a global distribution among a wide variety of marine fish species that serve as intermediate or paratenic hosts. The aim of this work was to study the occurrence of Anisakis spp. in the horse mackerel (Trachurus trachurus L.) from the North Atlantic Moroccan coasts. A fish sampling was conducted monthly during the August 2010 - July 2011 period. Samples were collected from the zone between Tangier and Larache. Morphological identification showed that the larvae found in this fish species correspond to Anisakis spp. Most third-stage (L3) larvae observed were found in the abdominal cavity and rarely in the muscle. The infection prevalence and intensity vary with the sampling period and fish size. A positive but non-significant correlation was found between infestation intensity and the fish host length ($R^2 = 0.25$). The highest mean intensities were observed for sizes > 28 cm during the March - June period.

Key words: Anisakis spp., Trachurus trachurus, Moroccan coasts.

Introduction

The Anisakidae belong to the Order of Ascaridida, parasitizing fish, mammals, birds and rarely reptiles (Hartwich, 1974). They have a global distribution among a wide variety of marine fish species that serve as intermediate or paratenic hosts (Koie *et al.*, 1995).

Anisakis spp. is one of the most damaging nematodes in terms of public health. Indeed, it is responsible of anisakiasis (zoonosis). After the consumption of raw or undercooked parasitized fish (Ishikura, 1989), the third-stage (L3) larvae invade the gastrointestinal mucosa and secrete proteins implicated in the anisakiasis pathogenesis (Matthews, 1982; Hotez *et al.*, 1994; Dziekonska-Rynko *et al.*, 2004). The penetration of this parasite in the tissue can induce allergic reactions with several clinical manifestations ranging from urticaria or angioedema to anaphylaxis (Kasuya et al., 1990; Audicana et al., 1995; Gonzalez-Quijada et al., 2005). Although they could not continue its development in humans, the larvae can become attached to the stomach or duodenum, causing digestive problems (pain, nausea, vomiting) which can be severe (Hartwich, 1974).

The horse mackerel [*Trachurus trachurus* Linné, 1758 (Quéro *et al.*, 2007) is among the principal hosts of L3 larvae of *Anisakis* spp., with a prevalence of 80 - 100% (Koie *et al.*, 1995). In the North of Morocco, horse mackerel (*T. trachurus*) presents one of

the fish species that are most commonly consumed. Horse mackerel acquires maturity at 3 years for males (20-22 cm) and around 4 or 5 years for females (26-30 cm) (Eaton, 1983; Eaton, 1989; Iversen et al., 1989). The juvenile ones feed mainly on copepods but also ostracods, mysids and fishes (gobies, run). In the adult stage, their diet consists mainly of fishes (anchovy, sprat, sardine, herring and sand lance), cephalopods and crustaceans (Quéro and Vayne, 1997). The analysis of stomach content of horse mackerel showed that it contains Crustacea (Euphausiacea, Mysidacea, Decapoda), Teleostei, Cephalopoda and Polychaeta. Euphausiacea were the most ingested (Santić et al., 2003). Euphausiids are intermediate hosts of Anisakis spp. (Adroher et al., 1996).

Despite the commercial and zoonotic risk of larval anisakid infections of fishes, their distribution among the Moroccan marine resources (teleosts) was not studied to our knowledge. The monitoring of Anisakis species prevalence to identify fishing areas most infected and the risked periods during the vear would contribute to establish effective control measures, useful for operators and consumers.

The present study focuses on surveying *Anisakis* spp., a nematode parasite, in *T. trachurus* caught in the Tangier-Larache coasts (NW of Morocco).

Materials and methods

A total of 205 specimens of *T*. *trachurus* were captured in the Tangier -Larache coasts (NW of Morocco), from August 2010 to July 2011.The sampling site was the wholesale fish market of Tangier. The individuals were taken randomly and represent various sizes, ranging from 18 to 38 cm.

Fish samples were measured, weighed and dissected by making an incision along the ventral line from the anus to the buccal aperture. The visceral cavity and digestive tract were examined. The muscle was removed from the spine and immersed in warm water for several minutes to facilitate the migration of the parasite to the outside of the fish.

After dissection of the individuals. encapsulated or free nematode larvae were collected from the visceral cavity, internal organs and musculature. Parasites were counted and, after being washed in a 0.9% NaCl, fixed in a 70% ethanol/glycerin solution (9/1), and cleared in glycerin solution 99% (Neveu-Lemaire, 1936). Then, they were observed in microscope. Larvae were identified according to previous works namely the structural features (Hartwich, 1974; Yoshinaga et al., 1987; Petter and Maillard, 1988; Ruiz-Valero et al., 1992; Ouiazon et al., 2008).

Parasitological indexes related to prevalence (P), intensity (I) and abundance (A) are calculated in accordance with Bush *et al.* (1997): P = [(number of infected fiches/number of fiches examined)*100]; I = (total number of parasites/total number of infected fish); A = (total number of parasites/total number of examined fish).

To compare mean prevalence, we used the Fisher's exact test. The bootstrapping 2-sample t- test is used to compare mean intensities and mean abundances. Comparisons based on normal theory such as Student's *t*-test or ANOVA are usually not applicable for parasites because of the skewness of their distribution. The statistical analysis was based on SPSS (Statistical Package for the Social Sciences) version 14.0 (Rózsa *et al.*, 2000).

Results

Localization of Anisakis spp. in T. Trachurus tissue

The parasite was found free or encysted in spiral form in different organs of mackerel: gonads, liver, intestine, viscera, mesentery and muscle. Larvae were frequently found in the abdominal cavity in visceral organs. Only a few ones were observed in the muscle (Figure 1).

Morphological identification

Anisakis spp. were identified on the basis of the morphological features. L3 larvae of Anisakis spp. are characterized by its whitish color. They have an oesophageal ventricle more or less developed. They are devoid of ventricular appendix and intestinal caecum. In the majority of *Anisakis* spp. larvae observed, the plan of ventricular-intestinal union is oblique and the posterior end is conical and ends with a mucro (Figure 2).

Effect of season variation on infection levels

The Anisakis spp. larvae were found with a prevalence of 100% in September-October 2010 and April-July 2011 periods. However in August 2010, the prevalence was relatively low (57.7%). This is due to the small size of individuals in the sample (23.63 cm, 108.1 g). The intensity of infection varies throughout the study period. It is more important in March and June 2011 and less important in August 2010 (Figure 3).

There was a high significant increase (p=0.01) in prevalence and intensity of infection with the increase

Figure 1. Anisakis spp. larvae in horse mackerel. A: L3 larvae showing esophageal ventricle. B: L3 larvae encapsulated on gonad. Legend: v (ventricle), \rightarrow (L3 Larvae).

Scale bar = 1 mm.







Figure 2. Morphological characteristics of L3 larvae of Anisakis spp. observed. A: Anterior end of Anisakis spp. L3 larvae. B: Tail of Anisakis spp. L3 Larvae. C: Ventricular part of Anisakis spp. (V). D: L3 larvae showing oblique junction ventricle between and intestine (arrowed). Legend: bt: boring tooth; i: intestine; m: mucro; v: ventricle.

Scale bar: 200µm.







Figure 4. Variation prevalence and intensity of infestation by L3 larvae of *Anisakis* spp. according to the fish length in horse mackerel.

of the total length of fish. The mean intensity of *Anisakis* spp. larvae was high in 31-38 cm fishes and low when the length fish is inferior to 26 cm. The mean prevalence was highest in fish exceeding 28 cm of size (96% for 29-30 cm class) and lowest in fish measuring less than 23 cm (Figure 4).

Discussion

The larvae and adults of *Anisakis* spp. are common parasites of a wide range of marine fish species (Smith and Wootten, 1978; Anderson, 1992; Williams and Jones 1994). *Anisakis* spp. occurs in fish at the larval stage and its infection of *T. trachurus* was previously reported in the Mediterranean and North Atlantic costs (Adroher *et al.*, 1996; Farjallah *et al.*, 2008; Mattiucci *et al.*, 2008). Horse mackerel may become infected with *Anisakis* larvae by

consuming euphausiids, which are intermediate hosts of these nematodes. It is a pelagic specie and eats small fishes and planktonic crustaceans (Muus and Dahlstrom, 1966; Smith, 1983).

When observed under the stereomicroscope and light microscope, L3 larvae of the nematode Anisakis spp. showed morphological characteristics similar to those mentioned by Rello-Yubero et al. (2004). These larvae are characterized by a cuticle with thin transversal striation. Anterior extremity is with a dorsal and two poorly developed ventro-lateral lips. We note the presence of six cephalic papillae, one pair in the dorsal lip and a pair in each ventro-lateral lip. Then, boring tooth is below the oral aperture. between the two ventro-lateral lips. The ventricle oesophagien is present, the ventricular appendix and the intestinal caecum are absent. There are also two nearly spherical rectal glands. The tail is conical, and the mucro is present. The morphological differentiation of Anisakis larvae was performed according to Smith and Wootten (1978) and Rello-Yubero et al. (2004) based on the two main morphological features: length of ventricular zone of the esophagus (shorter for A. physeteris) and the presence of a mucro at the extreme end of the tail in A. simplex s. str. Most of the larvae we observed belong to these two species, with predominance of A. simplex s. str. (ventricle esophageal longer than wide, presence of mucro, oblique union ventricle-intestine). In fact, a molecular study conducted by Farjallah et al. (2008) confirmed that in samples from the waters of Tangier, the highest frequencies for A. simplex s. str. was recorded. According to Mattiucci et al. (1997) and Mattiucci & Nascetti G (2006), A. simplex s. str. was the most prevalent species reported from the North-East Atlantic coasts, mainly from the North of Straits of Gibraltar. In accordance with Farjallah et al. (2008), 83% of the fish infected by A. simplex s. str. are pelagic and we think that the most species of Anisakis spp. we observed correspond probably to A. *simplex* s. str.

We observed that variations in the length of fish sampled each month correlated with variations in *Anisakis* spp. prevalence throughout the year. Furthermore, *Anisakis* spp. mean prevalence was higher in the second half of the year. According to Smith and Wootten (1978), seasonal variations in infection levels are probably due to changes in the population of infected euphausiids in the zooplankton.

The highest levels of infection intensity were found in March and June, with a mean intensity of 34 and 48.6 in the same order, similarly to Adroher *et al.* (1996).

We also observed that parasite mean intensity varied directly with the length (age) of horse mackerel.

Adroher *et al.* (1996) studied variations in the prevalence of *Anisakis* spp. according to the host size in horse mackerel. Similarly to our results, they found that fishes measuring more than 30 cm in length were infected more frequently than those measuring less than 23 cm. Most larvae were observed in body cavity, and only some of them were found in muscles. These results are similar to those reported by Huang (1988) and Adroher *et al.* (1996) who reported that most larvae occurred in the body cavity (61.2%).

Acknowledgments

The authors thank Benchoucha Said and Rharrabe Kacem for the manuscript revision.

References

Adroher FJ, Valero A, Ruiz-Valero J, Iglesias L (1996) Larval anisakids (Nematoda: Ascaridoidea) in horse mackerel (*Trachurus trachurus*) from the fish market in Granada (Spain). *Parasitol Res* 82: 253-256.

Anderson RC (1992) Nematode parasites of vertebrates: their development and transmission. CAB International Publishing, Wallingford, 650.

Audicana M, Ferna'ndez de Corres L, Muňoz D, Fernandez E, Navarro JA, del Pozo MD (1995) Recurrent anaphylaxis caused by *Anisakis simplex* parasitizing fish. *J. Allergy Clin. Immunol* **6** : 558– 560.

Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997) Parasitology meets ecology on its own terms: Margolis et al. Revisited *Journal of Parasitology* **83**: 575-583.

Dziekonska-Rynko J, Rokicki J, Jablonowski Z (2004) Efects of 3rd stage Anisakis simplex larvae on digestive tract protease activity of guinea pigs 24 and 48 hours after infection. *Helminthologia* **1**: 1–24.

Eaton DR (1983) Scad in the north-east Atlantic. Lab. Leafl., MAFF Direct. Fish. Res., Lowestoft, **56** : 20.

Eaton DR (1989) Spawning-stock biomass of scad (*Trachurus trachurus* L.) to the west of the British Isles, as indicated by egg surveys. J. Con. int. Explor. Mer **45**: 231-247.

Farjallah S, Busi B, Ould Mahjoub M, Ben Slimane B, Paggi L, Said K, D'Amelio S (2008) Molecular characterization of larval anisakid nematodes from marine fishes off the Moroccan and Mauritanian coasts. *Parasitology International* **57:** 430-436.

Gonzalez-Quijada S, Gonzalez Escudero R, Arias Garcia L, Gil Martin AR, Vicente Serrano J, Corral-Fernandez E (2005) Anisakiasis gastrointestinal manifestations: description of 42 cases. *Rev. Clin. Esp* **205:** 311–315.

Hartwich G (1974) CIH keys to the nematode parasites of vertebrates. In: Anderson RC, Chabaud AG & Willmott S (Eds.), Keys to genera of the Ascaridoidea. CAB, Slough, **2**: 15.

Hotez P, Cappello M, Hawdon J, Beckers C, Sakanari J (1994) Hyaluronidases of the gastrointestinal invasive nematodes Ancylostoma caninum and *Anisakis* simplex: possible functions in the pathogenesis of human zoonoses *J. Infect. Dis* **70**: 918–926.

Huang W 1988 Anisakides et anisakidoses humaines. Deuxième partie: enquête sur les anisakidés de poissons commerciaux du marché parisien. Ann Parasitol Hum Comp 63:197–208.

Ishikura H (1989) General survey of *Anisakis* and Anisakiasis in Japan. In: Ishikura H & Namiki M (Eds.), Gastric Anisakiasis in Japan: Epidemiology, diagnosis treatment. *Springer-Verlag*, Tokyo, 3-11.

Iversen SA, Eltink A, Kirkegaard E, Skagen DW (1989) The egg production and spawning stock size of the North Sea mackerel and horse mackerel stocks in 1988. Int. Coun. Explor. Sea CM 1989/H **16**: 22.

Kasuya S, Hamano H, Izumi S (1990) Mackerel-induced urticaria and *Anisakis. Lancet* **35**: 65.

Koie M, Berland B, Burt MDB (1995) Developement to third-stage larva occurs in the eggs of *Anisakis simplex* and *Pseudotarranova decipiens* (Nematoda, Ascaridoidea, Anisakidae). *Can J Fish Aquat Sci* **52**:134-139.

Matthews BE (1982) Behaviour and enzyme release by *Anisakis*. sp larvae (Nematoda: Ascaridida). *J. Helminthol* **6:** 77–183.

Mattiucci S, Farina V, Campbell N, MacKenzie K, Ramos P, Pinto AL, Abaunza P, Nascetti G (2008) *Anisakis* spp. larvae (Nematoda: Anisakidae) from Atlantic horse mackerel: Their genetic identification and use as biological tags for host stock characterization. *Fisheries Research* **89**: 146-151.

Mattiucci S, Nascetti G (2006) Molecular systematics, phylogeny and ecology of anisakid nematodes of the genus *Anisakis* Dujardin, 1845: *an update*. *Parasite* **13**: 99-113.

Mattiucci S, Nascetti G, Cianchi R, Paggi L, Arduino P, Margolis L, Brattey J, Webb SC, D'Amelio S, Orecchia P, Bullini L (1997) Genetic and ecological data on the *Anisakis simplex* complex with evidence for a new species (Nematoda, Ascaridoidea, Anisakidae). *J Parasitol* **83:** 401-416.

Muus BJ, Dahlstrom P (1966) Guide des poissons de mer et pêche. Delachaux & Niestlé, Neuchâtel, Switzerland.

Neveu-Lemaire M (1936) Traité d'helminthologie médicale et vétérinaire. Vigor Frères, Paris, 1515.

Petter AJ, Maillard C (1988) Larves d'ascarides parasites de poissons en Méditerranée occidentale. *Bulletin du Muséum National d'Histoire Naturelle*, Paris. **10**: 347-369.

Quéro GC, Spitz J, Vayen JJ (2007) Faune française de l'atlantique : poissons Carangidés. *Annales de la société des sciences naturelles Charente-Maritime* **7**: 709-722.

Quéro JC and Vayne JJ (1997) Les poissons de mer des pêches françaises. Ifremer, Delachaux & Niestlé (Eds.), 304.

Quiazon AKM, Yoshinaga T, Ogawa K, Yukami R (2008) Morphological differences between larvae and in vitrocultured adults of *Anisakis simplex* (sensu stricto) and *Anisakis pegreffii* (Nematoda: Anisakidae). Parasitology International **57:** 483-489.

Rello-Yubero FJ, Adroher-Auroux FJ, Valero-López A (2004) Anisákidos parásitos de peces comerciales. Riesgos asociados a la salud pública. Anales de la Real Academia de ciencias Veterinarias de Andalucía Oriental **17**: 173-197. Rózsa S, Reiczigel J, Majoros G (2000) Quantifying parasites in samples of hosts. *J. Parasitol* **86** (2): 282-232.

Ruiz-Valero J, Valero A, Adroher FJ, Ortega JE (1992) Presencia de ascáridos en peces comerciales de frecuente consumo en Granada. In: Hernández-Rodríguez S (ed.), "In memoriam" al Profesor Doctor DF de P Martínez Gómez. Universidad de Córdoba, Spain: 335–349.

Santić M, Jardas I, Pallaoro A (2003) Feeding habits of Mediterranean horse mackerel *Trachurus mediterraneus* (Carangidae) in the central Adriatic sea. *Cybium* **27(4)**: 247-253.

Smith JW (1983) Anisakis simplex (Rudolphi, 1809, det. Krabbe, 1878) (Nematoda: Ascaridoidea): morphology and morphometry of larvae from euphausiids and fish, and a review of the life-history and ecology. J Helminthol **57:**205–224

Smith JW, Wootten R (1978) Anisakis and anisakiasis. Advances in Parasitology **16:** 93-163.

Williams HH, Jones A (1994) Parasitic worms of fish. Taylor & Francis, CRC Press 1 edition, London, 593.

Yoshinaga T, Ogawa K, Wakabayashi H (1987) Experimental life cycle of *Hysterothylacium aduncum* (Nematoda: Anisakidae) in fresh water. *Fish Pathol* **22:** 243-251.